

available at www.sciencedirect.comjournal homepage: www.ejconline.com

Wnt1 overexpression promotes tumour progression in non-small cell lung cancer

Cheng-Long Huang^{a,*}, Dage Liu^a, Shinya Ishikawa^a, Takashi Nakashima^a,
Nariyasu Nakashima^a, Hiroyasu Yokomise^a, Kyuichi Kadota^b, Masaki Ueno^c

^aDepartment of General Thoracic Surgery, Breast and Endocrinological Surgery, Faculty of Medicine, Kagawa University, 1750-1, Miki-cho, Kita-gun, Kagawa 761-0793, Japan

^bDepartment of Diagnostic Pathology, Faculty of Medicine, Kagawa University, 1750-1, Miki-cho, Kita-gun, Kagawa 761-0793, Japan

^cDepartment of Pathology and Host Defense, Faculty of Medicine, Kagawa University, 1750-1, Miki-cho, Kita-gun, Kagawa 761-0793, Japan

ARTICLE INFO

Article history:

Received 18 June 2008

Received in revised form 31 July 2008

Accepted 1 August 2008

Available online 13 September 2008

Keywords:

Wnt1

c-Myc

Cyclin D1

VEGF-A

MMP-7

Lung cancer

ABSTRACT

Background: The Wnt gene family is involved in embryogenesis and tumorigenesis. We investigated the clinical significance of Wnt1 expression in non-small cell lung cancer (NSCLC).

Method: We studied 216 NSCLC patients. Immunohistochemistry was performed to investigate the Wnt1 expression in relation to the expression of β -catenin and Wnt-targets, including c-Myc, Cyclin D1, VEGF-A and MMP-7. The Ki-67 proliferation index and the intratumoural microvessel density (IMD) were also evaluated.

Results: The ratio of tumours with an aberrant β -catenin expression was significantly higher in Wnt1-positive tumours than in Wnt1-negative tumours ($p < 0.0001$). The Wnt1 expression significantly correlated with the expression of c-Myc ($p < 0.0001$), Cyclin D1 ($p < 0.0001$), VEGF-A ($p = 0.0160$), MMP-7 ($p < 0.0001$), the Ki-67 index ($p = 0.0048$) and the IMD ($p = 0.0267$). Furthermore, the Wnt1 status was a significant prognostic factor for NSCLC patients ($p = 0.0127$).

Conclusions: The Wnt1 overexpression is associated with the expression of tumour-associated Wnt-targets, tumour proliferation, angiogenesis and a poor prognosis in NSCLCs.

© 2008 Elsevier Ltd. All rights reserved.

1. Introduction

Non-small cell lung cancer (NSCLC) is one of the most common human cancers with a poor prognosis. However, recent molecular biology studies have revealed that many molecules affect the various biological behaviours of malignant tumours. It is therefore considered important to clarify these tumour biology mechanisms in order to improve the clinical outcome of NSCLC patients.¹ The Wnt gene family encodes the multi-functional signalling glycoproteins that are in-

involved in the regulation of a wide variety of normal and pathological processes, including embryogenesis, differentiation and tumorigenesis.^{2,3} The Wnt genes have been classified into functional groups with separate downstream signalling pathways.⁴ Amongst them, Wnt1 stimulates the canonical Wnt/ β -catenin signalling pathway, which leads to changes in cell fate and/or cell transformation.⁵ The canonical Wnt/ β -catenin signalling pathway regulates the transcription of the Wnt-target genes with TCF/LEF1 motifs.⁶ The Wnt-target genes include various molecules associated with tumorigenesis.^{7–10}

* Corresponding author. Tel.: +81 87 891 2191; fax: +81 87 891 2192.

E-mail address: chuang@kms.ac.jp (C.-L. Huang).

0959-8049/\$ - see front matter © 2008 Elsevier Ltd. All rights reserved.

doi:10.1016/j.ejca.2008.08.004

As a result, the intratumoural Wnt1 expression could affect various biological functions through these Wnt-target genes. In fact, previous clinical studies have demonstrated that the Wnt1 expression is frequently upregulated in many human cancers.^{11,12}

Therefore, the Wnt1 overexpression could affect tumour biology during tumour progression and understanding the biological mechanisms of the Wnt1 expression could lead to the development of a new treatment for cancer patients. Therefore, in order to clarify the biological significance of the Wnt1 expression in NSCLCs, we investigated the intratumoural Wnt1 expression in relation to its target molecules, including c-Myc,⁷ Cyclin D1,⁸ vascular endothelial growth factor-A (VEGF-A)⁹ and matrix metalloproteinase-7 (MMP-7).¹⁰ In addition, we also evaluated the tumour proliferation rate using the Ki-67 index, and tumour angiogenesis using CD34 staining.¹

2. Materials and methods

2.1. Clinical characteristics of the patients

From January 1996 to December 2002, consecutive NSCLC patients who underwent surgery at the Department of General Thoracic Surgery, Breast and Endocrinological Surgery of Kagawa University were studied. This study was approved by the institutional review board of Kagawa University (14-7, a clinical study of biological markers in non-small cell lung cancers), and a signed informed consent was obtained from each patient. Tumour-node-metastasis (TNM) staging designations were made according to the postsurgical pathological international staging system. The lymph node status was pathologically evaluated using specimens resected by either thoracotomy or mediastinoscopy. In total, 216 patients with lung cancer up to stage IIIB, which included 123 patients with adenocarcinomas, 83 patients with squamous cell carcinomas and 10 patients with large cell carcinomas, were investigated (Table 1). The patients' clinical records and histopathological diagnoses were fully documented. This report includes follow-up data as of October 2007. Systemic chemotherapy using mitomycin/vinblastin/cisplatin or carboplatin/paclitaxel was performed in all patients with stages II–III NSCLCs: neoadjuvant chemotherapy in 43 patients and postoperative adjuvant chemotherapy in 50 patients. Radiation therapy was performed in 26 patients: twelve patients with T3 or T4 status and 14 patients with mediastinal lymph node metastases.

2.2. Immunohistochemical assays

We performed immunohistochemical studies to evaluate the intratumoural expression of Wnt1, β -catenin, c-Myc, Cyclin D1, VEGF-A and MMP-7, tumour proliferation rate using the Ki-67 index and tumour angiogenesis using CD34 staining. The following antibodies were used, along with isotype antibodies as negative controls: a rabbit polyclonal antibody for Wnt1 (H-89, Santa Cruz Biotechnology, Santa Cruz, CA, USA) diluted at 1:200, a mouse monoclonal antibody for β -catenin (C19220, Transduction Laboratories, Lexington, KY) diluted at 1:200, a mouse monoclonal antibody for c-Myc (9E10, Santa

Table 1 – Demographic and clinical characteristics of patients

| Characteristics | Number of patients | Percent |
|---|--------------------|---------|
| Total number of patients | 216 | 100 |
| <i>Age, years</i> | | |
| Median | 68 | |
| Range | 38–82 | |
| <i>Gender</i> | | |
| Male | 154 | 71.3 |
| Female | 62 | 28.7 |
| <i>Smoking status</i> | | |
| Non-smoker | 75 | 34.7 |
| Smoker | 141 | 65.3 |
| <i>Smoking pack-years</i> | | |
| Median | 36 | |
| Range | 0–150 | |
| <i>ECOG performance status</i> | | |
| 0 | 139 | 64.3 |
| 1 | 52 | 24.1 |
| 2 | 25 | 11.6 |
| <i>Histology</i> | | |
| Adenocarcinoma | 123 | 56.9 |
| Squamous cell carcinoma | 83 | 38.4 |
| Large cell carcinoma | 10 | 4.7 |
| <i>Pathological stages</i> | | |
| I | 123 | 56.9 |
| II | 27 | 12.5 |
| III | 66 | 30.6 |
| <i>Method of surgical resection</i> | | |
| Pneumonectomy | 20 | 9.3 |
| Lobectomy | 179 | 82.8 |
| Segmentectomy | 9 | 4.2 |
| Wedge resection | 8 | 3.7 |
| <i>Chemotherapy</i> | | |
| Neoadjuvant therapy | 43 | 19.9 |
| Postoperative adjuvant therapy | 50 | 23.2 |
| <i>Radiotherapy</i> | | |
| Radiotherapy | 26 | 12.0 |
| Abbreviation: ECOG, Eastern Cooperative Oncology Group. | | |

Cruz) diluted at 1:100, a mouse monoclonal antibody for Cyclin D1 (DSC-6, DAKO, Glostrup, Denmark) diluted at 1:200, a rabbit polyclonal antibody for VEGF-A (A-20, Santa Cruz) diluted at 1:200, a rabbit polyclonal antibody for MMP-7 (AB19135, Chemicon, Temecula, CA, USA) diluted at 1:300, a mouse monoclonal antibody for the Ki-67 antigen (MIB-1, DAKO) diluted at 1:40 and a mouse monoclonal antibody for CD34 (NU-4A1, Nichirei Corporation, Tokyo, Japan) diluted at 1:10.

Formalin-fixed paraffin-embedded tissue was cut into 4- μ m sections and mounted on poly-L-lysine-coated slides. Sections were deparaffinised and rehydrated. The slides were then heated in a microwave for 10 min in a 10- μ mol/l citrate buffer solution at pH 6.0, and cooled to room temperature. After quenching the endogenous peroxidase activity with 0.3% H₂O₂ (in absolute methanol) for 30 min, the sections were treated for 2 h at room temperature with 5% bovine serum albumin to block non-specific staining. Duplicate sections were incubated overnight with the primary specific antibodies.

Slides were then incubated for 1 h with biotinylated anti-rabbit IgG (Vector Laboratories Inc., Burlingame, CA) for Wnt1, VEGF-A and MMP-7 and biotinylated anti-mouse IgG (Vector Laboratories Inc.) for β -catenin, c-Myc, Cyclin D1, Ki-67 and CD34. The sections were incubated with the avidin-biotin-peroxidase complex (Vector Laboratories Inc.) for 1 h, and antibody binding was visualised with 3,3'-diaminobenzidine tetrahydrochloride. Lastly, the sections were lightly counterstained with Mayer's haematoxylin. Sections of lung tumours known to express Wnt1, c-Myc, Cyclin D1, VEGF-A or MMP-7 were used as positive controls for the immunohistochemical staining, respectively.

All the immunostained sections were reviewed by two authors (Huang and Ueno) who had no knowledge of the patients' clinical status. Cases with discrepancies were jointly re-evaluated, and a consensus was reached. For the expression of Wnt1, c-Myc, Cyclin D1, VEGF-A and MMP-7, five areas were selected at random and scored in cases with multiple areas of low intensity. Also, one random field was selected in sections where all staining appeared intense. At least 200 cells were scored per $\times 40$ field about tumour cells. Regarding the β -catenin expression, we used the classification of staining patterns as reported previously:¹³ (1) a membranous pattern, if immunoreactivity was present solely at the cell membranes; (2) a membranous-cytoplasmic pattern, if immunoreactivity was also present in the cytoplasm; (3) a cytoplasmic pattern, if immunoreactivity was chiefly present in the cytoplasm and in less than 20% of the nuclei and (4) a cytoplasmic-nuclear pattern, if immunoreactivity was present in the cytoplasm and concomitantly in more than 20% of the nuclei. The percentage of carcinoma cells with a positive staining for Ki-67 in a given specimen was scored as the Ki-67 proliferation index. For microvessel quantification, the three most highly vascularised areas detected by CD34 immunostaining were initially selected under the $\times 40$ field, and a $\times 200$ field (0.785 mm² per field) was used to count vessels in each of these areas. The average of three $\times 200$ field counts was recorded as the intratumoural microvessel density (IMD).¹

2.3. Statistical analysis

Because the distributions of seven values, including the percentages of Wnt1-positive tumour cells ($p = 0.2128$), c-Myc-positive tumour cells ($p = 0.1390$), Cyclin D1-positive tumour cells ($p = 0.1390$), VEGF-A-positive tumour cells ($p = 0.0531$), MMP-7-positive tumour cells ($p = 0.0875$), the Ki-67 proliferation index ($p = 0.0875$) and the IMD ($p = 0.1727$), showed normal distributions (Kolmogorov-Smirnov analysis), the statistical significances regarding these values were assessed by the t-test, ANOVA with Bonferroni/Dunn test or Pearson's correlation coefficient. The sample was classified as a Wnt1-positive tumour when the percentage of Wnt1-positive tumour cells was $>50\%$ because of the most significance in relation to the Ki-67 proliferation index, which is the same as reported previously.¹⁴

Overall survival was defined as the time from the treatment initiation (surgical resection, chemotherapy or radiation) to the date of death from any cause. The Kaplan-Meier method was used to estimate the probability of overall sur-

vival as a function of time, and differences in the survival of subgroups of patients were compared by using Mantel's log-rank test. A multivariate analysis was performed using the Cox regression model to study the effects of different variables on survival. All p values were based on the two-sided statistical analysis, and a p value of <0.05 was considered to indicate statistical significance.

3. Results

3.1. Wnt1 expression in NSCLCs

The Wnt1 expression appeared in the form of a cytoplasmic staining pattern (Fig. 1A). The Wnt1 expression was low in the normal alveolar epithelium. In contrast, regarding the intratumoural Wnt1 expression, the percentage of Wnt1-positive tumour cells varied greatly amongst the 216 NSCLCs (median, 45.5%; mean \pm SD, $46.7 \pm 27.9\%$). One hundred and six carcinomas (49.1%) were Wnt1-positive (Table 2). No significant difference in the Wnt1 status was observed according to the tumour histology, tumour status, nodal status, pathological stage or tumour differentiation.

3.2. The β -catenin expression in relation to the Wnt1 status

The intratumoural β -catenin expression exhibited four staining patterns (Fig. 1B and H).¹³ Amongst 106 Wnt1-positive tumours, 8 carcinomas (7.5%) had a membranous pattern, 21 carcinomas (19.8%) had a membranous-cytoplasmic pattern, 37 carcinomas (34.9%) had a cytoplasmic pattern and 40 carcinomas (37.7%) had a cytoplasmic-nuclear pattern. Amongst 110 Wnt1-negative carcinomas, 37 carcinomas (33.6%) had a membranous pattern, 45 carcinomas (40.9%) had a membranous-cytoplasmic pattern, 11 carcinomas (10.0%) had a cytoplasmic pattern and 17 carcinomas (15.5%) had a cytoplasmic-nuclear pattern. The ratio of tumours with a cytoplasmic-nuclear pattern or a cytoplasmic pattern was significantly higher in Wnt1-positive tumours than in Wnt1-negative tumours (72.6% versus 25.5%; $p < 0.0001$).

3.3. The c-Myc expression in relation to the Wnt1 status

In order to investigate the biological functions of the Wnt1 expression in NSCLC, we studied four tumour-associated Wnt1-targets such as c-Myc, Cyclin D1, VEGF-A and MMP-7. Regarding the intratumoural c-Myc expression, the percentage of c-Myc-positive tumour cells varied greatly amongst the 216 NSCLCs (median, 39.0%; mean \pm SD, $39.5 \pm 28.8\%$; Fig. 1C). Furthermore, the percentage of Wnt1-positive tumour cells significantly correlated with the percentage of c-Myc-positive tumour cells ($r = 0.376$; $p < 0.0001$; Fig. 2A). The percentage of c-Myc-positive tumour cells was significantly higher in Wnt1-positive tumours than in Wnt1-negative tumours ($50.2 \pm 26.8\%$ versus $29.2 \pm 27.0\%$; $p < 0.0001$; Fig. 2B). Regarding the clinical significance of c-Myc status, the percentage of c-Myc-positive tumour cells significantly correlated with the Ki-67 proliferation index ($r = 0.328$; $p < 0.0001$).

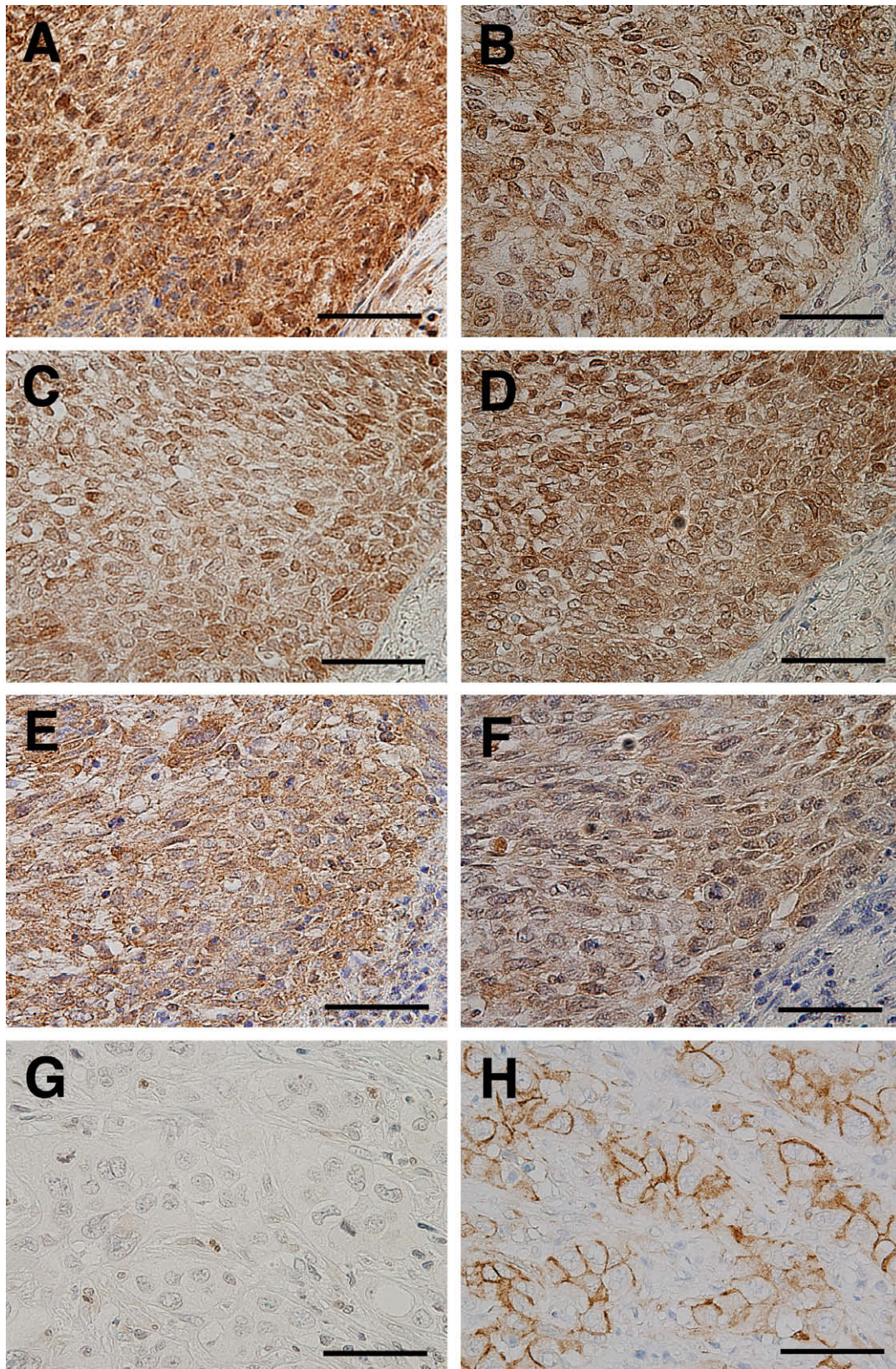


Fig. 1 – Immunostaining of lung cancers. A carcinoma with the positive expression of Wnt1 (A), cytoplasmic-nuclear expression of β -catenin (B), positive expression of c-Myc (C), positive expression of Cyclin D1 (D), positive expression of VEGF-A (E) and positive expression of MMP-7 (F). A carcinoma with negative expression of Wnt1 (G) and membranous expression of β -catenin (H). Bar, 50 μ m.

Table 2 – Distribution of Wnt1 status in NSCLC patients according to clinicopathological characteristics

| Characteristics | Number of patients | Wnt1 status | | p-Value |
|---------------------------------|--------------------|-------------|------------|---------|
| | | Negative | Positive | |
| Smoking | | | | |
| Non-smoker | 75 | 35 | 40 | 0.3611 |
| Smoker | 141 | 75 | 66 | |
| Tumour status | | | | |
| T1 | 87 | 45 | 42 | 0.3212 |
| T2 | 72 | 39 | 33 | |
| T3 | 18 | 11 | 7 | |
| T4 | 39 | 15 | 24 | |
| Nodal status | | | | |
| N0 | 153 | 81 | 72 | 0.3559 |
| N1, N2, N3 | 63 | 29 | 34 | |
| Pathological stage | | | | |
| I | 123 | 65 | 58 | 0.3294 |
| II | 27 | 16 | 11 | |
| III | 66 | 29 | 37 | |
| Differentiation | | | | |
| Well | 75 | 44 | 31 | 0.2221 |
| Moderately | 78 | 38 | 40 | |
| Poorly | 63 | 28 | 35 | |
| Histology | | | | |
| Adenocarcinoma | 123 | 67 | 56 | 0.0832 |
| Squamous cell carcinoma | 83 | 35 | 48 | |
| Large cell carcinoma | 10 | 8 | 2 | |
| Total number of patients | 216 | 110 | 106 | |

3.4. The Cyclin D1 expression in relation to the Wnt1 status

Regarding the intratumoural Cyclin D1 expression, the percentage of Cyclin D1-positive tumour cells varied greatly amongst the 216 NSCLCs (median, 38.5%; mean \pm SD, $39.9 \pm 26.2\%$; Fig. 1D). The percentage of Wnt1-positive tumour cells also significantly correlated with the percentage of Cyclin D1-positive tumour cells ($r = 0.410$; $p < 0.0001$; Fig. 2C). The percentage of Cyclin D1-positive tumour cells was significantly higher in Wnt1-positive tumours than in Wnt1-negative tumours ($50.3 \pm 26.3\%$ versus $29.8 \pm 22.0\%$; $p < 0.0001$; Fig. 2D). Regarding the clinical significance of Cyclin D1 status, the percentage of Cyclin D1-positive tumour cells significantly correlated with the Ki-67 proliferation index ($r = 0.265$; $p < 0.0001$).

3.5. The VEGF-A expression in relation to the Wnt1 status

Regarding the intratumoural VEGF-A expression, the percentage of VEGF-A-positive tumour cells varied greatly amongst NSCLCs (median, 33.0%; mean \pm SD, $35.5 \pm 29.8\%$; Fig. 1E). The percentage of Wnt1-positive tumour cells significantly correlated with the percentage of VEGF-A-positive tumour cells ($r = 0.164$, $p = 0.0160$). The percentage of VEGF-A-positive tumour cells was significantly higher in Wnt1-positive tumours than in Wnt1-negative tumours ($40.7 \pm 31.8\%$ versus $30.5 \pm 27.0\%$; $p = 0.0119$; Fig. 2E). Regarding the clinical significance of VEGF-A status, the percentage of VEGF-A-positive tumour cells significantly correlated with the IMD ($r = 0.136$; $p = 0.0452$).

3.6. The MMP-7 expression in relation to the Wnt1 status

The percentage of MMP-7-positive tumour cells also varied greatly amongst the 216 NSCLCs (median, 48.0%; mean \pm SD, $45.9 \pm 30.2\%$; Fig. 1F). The percentage of Wnt1-positive tumour cells significantly correlated with the percentage of MMP-7-positive tumour cells ($r = 0.383$; $p < 0.0001$). The percentage of MMP-7-positive tumour cells was significantly higher in Wnt1-positive tumours than in Wnt1-negative tumours ($56.7 \pm 27.5\%$ versus $35.7 \pm 29.3\%$; $p < 0.0001$; Fig. 2F).

3.7. The clinical significance of the intratumoural Wnt1 status in NSCLC

Regarding tumour proliferation, the percentage of Wnt1-positive tumour cells significantly correlated with the Ki-67 proliferation index ($r = 0.191$; $p = 0.0048$). The Ki-67 proliferation index was significantly higher in Wnt1-positive tumours than in Wnt1-negative tumours ($53.2 \pm 29.5\%$ versus $38.8 \pm 28.4\%$; $p = 0.0003$; Fig. 2G).

Concerning tumour angiogenesis, the percentage of Wnt1-positive tumour cells also significantly correlated with the IMD ($r = 0.151$; $p = 0.0267$). The IMD was significantly higher in Wnt1-positive tumours than in Wnt1-negative tumours (113.1 ± 66.3 versus 96.8 ± 47.8 ; $p = 0.0383$; Fig. 2H).

Regarding the survival, the 5-year survival was 42.6% in patients with Wnt1-positive NSCLCs and 66.5% in patients with Wnt1-negative NSCLCs. The overall survival was significantly lower in patients with Wnt1-positive NSCLCs than in patients with Wnt1-negative NSCLCs ($p = 0.0013$; Fig. 3). A

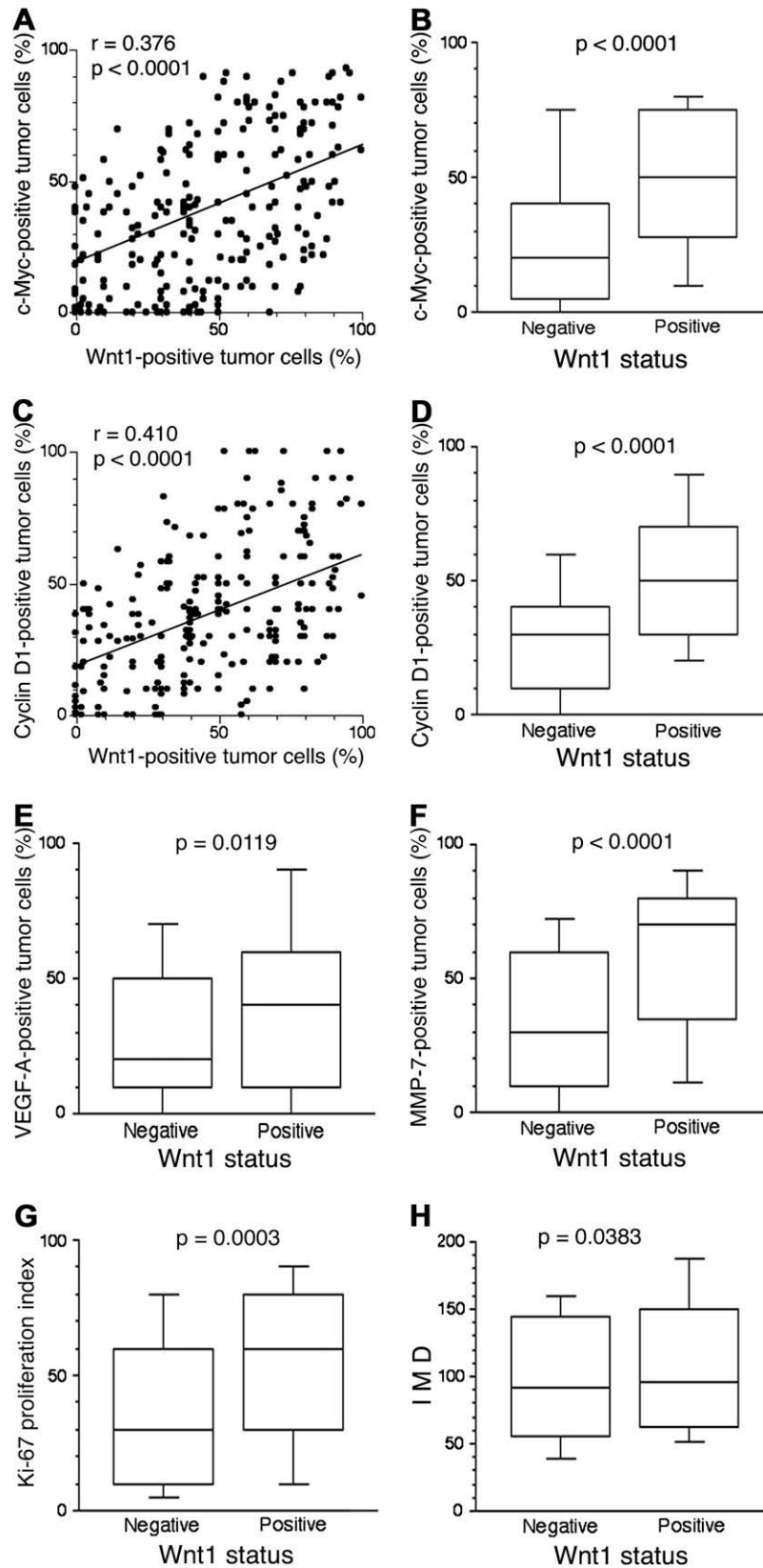


Fig. 2 – (A) The relationship between Wnt1 expression and c-Myc expression, **(B)** c-Myc expression in relation to the Wnt1 status, **(C)** The relationship between Wnt1 expression and Cyclin D1 expression, **(D)** cyclin D1 expression in relation to the Wnt1 status, **(E)** VEGF-A expression in relation to the Wnt1 status, **(F)** MMP-7 expression in relation to the Wnt1 status, **(G)** Ki-67 proliferation index in relation to the Wnt1 status and **(H)** IMD in relation to the Wnt1 status.

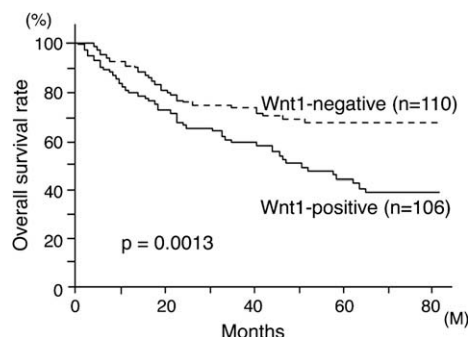


Fig. 3 – Overall survival of NSCLC patients in relation to the Wnt1 status.

multivariate analysis demonstrated that the Wnt1 status (hazard ratio 1.784; $p = 0.0127$), the tumour status (hazard ratio 1.280; $p = 0.0113$) and the nodal status (hazard ratio 1.479; $p = 0.0013$) were significant prognostic factors for NSCLC patients (Table 3).

4. Discussion

The Wnt genes encode secreted proteins with multi-directional biological functions via autocrine or paracrine routes.² They are involved in the regulation of a wide variety of normal

and pathological processes, including tumourigenesis.^{2,3} Amongst them, Wnt1 is one of the novel members stimulating the canonical Wnt/ β -catenin signalling pathway.⁵ The activation of the canonical Wnt signalling pathway leads to the stabilisation of β -catenin, subsequently regulating the transcription of many Wnt-target genes with TCF/LEF1 motifs.⁶ The Wnt-target genes include various molecules associated with tumourigenesis.^{7–10} Therefore, the Wnt1 overexpression could affect the tumour biology during tumour progression.

This clinical study in NSCLC has demonstrated that the intratumoural Wnt1 expression correlates with various tumour-associated Wnt-targets, including c-Myc,⁷ Cyclin D1,⁸ VEGF-A⁹ and MMP-7.¹⁰ As a result, the intratumoural Wnt1 expression was associated with the tumour proliferation, angiogenesis and a poor prognosis in NSCLC patients.

At first, β -catenin is a pivotal component of the canonical Wnt signalling pathway and E-cadherin-associated homotypic cell adhesion.¹⁵ The β -catenin expression is membranous in normal epithelium. In contrast, β -catenin demonstrated four expression patterns in tumour cells. The β -catenin expression in the cytoplasm and/or nuclear could be considered to be an indication of its aberrant expression.¹³ This study demonstrated that the aberrant expression of β -catenin was higher in Wnt1-positive NSCLCs than in Wnt1-negative NSCLCs. Although 28 Wnt1-negative carcinomas (12.8%) had the aberrant expression of β -catenin in this study,

Table 3 – Multivariate regression analysis in predicting the overall survival of NSCLC patients

| Variables | Assigned score | Hazard ratio | 95% CI | p-Value |
|-----------------|----------------|--------------|---------------|---------|
| Wnt1 status | | | | |
| Negative | 0 | | | |
| Positive | 1 | 1.784 | (1.132–2.813) | 0.0127 |
| Tumour status | | | | |
| T1 | 1 | | | |
| T2 | 2 | 1.280 | (1.058–1.550) | 0.0113 |
| T3 | 3 | | | |
| T4 | 4 | | | |
| Nodal status | | | | |
| N0 | 0 | | | |
| N1 | 1 | 1.479 | (1.166–1.876) | 0.0013 |
| N2 | 2 | | | |
| N3 | 3 | | | |
| Age | | | | |
| <60 | 0 | | | |
| ≥60 | 1 | 1.003 | (0.597–1.687) | 0.9897 |
| Gender | | | | |
| Female | 0 | | | |
| Male | 1 | 1.815 | (0.908–3.627) | 0.0915 |
| Smoking | | | | |
| Non-smoker | 0 | | | |
| Smoker | 1 | 0.798 | (0.422–1.507) | 0.4861 |
| Differentiation | | | | |
| Well | 0 | | | |
| Moderately | 1 | 1.195 | (0.890–1.604) | 0.2355 |
| Poorly | 2 | | | |

Abbreviation: CI, confidence interval.

this might be partly due to other members of the Wnt family, such as Wnt2b.

Regarding targets of the canonical Wnt/ β -catenin signalling pathway, the present study demonstrated the intratumoural Wnt1 expression to correlate with the expression of a transcription factor c-Myc.⁷ c-Myc is involved in growth control and cell cycle progression by stimulating and repressing the expression of cell cycle regulators.¹⁶ Therefore, c-Myc has pro-mutagenic effects in cancer cell lines. In fact, previous clinical studies have revealed that the c-Myc overexpression is associated with the malignant phenotype in various human cancers.^{17,18} This clinical study also demonstrated that the intratumoural c-Myc expression correlated with the tumour proliferation of NSCLC patients. Furthermore, amongst NSCLC patients we studied, the intratumoural c-Myc expression was significantly associated with the patient survival (data not shown).

Cyclin D1 is another target of the canonical Wnt/ β -catenin signalling pathway.⁸ This study revealed the intratumoural Wnt1 expression to correlate with the Cyclin D1 expression in NSCLCs. Although the Cyclin D1 gene is frequently amplified in lung carcinomas,¹⁹ this study identified another mechanism of the Cyclin D1 overexpression in NSCLC. Cyclin D1 regulates the G1-to-S phase transition.²⁰ In fact, its overexpression has been reported to be associated with the tumour proliferation,²¹ as also observed in this study.

On the other hand, angiogenesis is essential for tumour growth and metastasis. VEGF-A is a potent and widely distributed angiogenic peptide.²² Many clinical studies revealed the intratumoural VEGF-A expression to be associated with the tumour angiogenesis and a poor prognosis in cancer patients.^{1,23} In fact, amongst NSCLC patients we studied, the intratumoural VEGF-A expression was significantly associated with the patient survival (data not shown). In addition, Bevacizumab (Avastin; Genentech, Inc, South San Francisco, CA), a recombinant humanised version of the murine antihuman VEGF-A monoclonal antibody, has been developed for the clinical treatment of cancer patients, including NSCLC patients.²⁴ The VEGF-A is also a target of the canonical Wnt/ β -catenin signalling pathway.⁹ This study revealed that the Wnt1 expression correlated with the intratumoural VEGF-A expression.

MMP-7 is also one of the targets of the canonical Wnt/ β -catenin signalling pathway.¹⁰ This study demonstrated the intratumoural Wnt1 expression to correlate with the MMP-7 expression in NSCLCs. Although the biological functions of MMP-7 are still not clearly understood, previous clinical studies have shown an overexpression of MMP-7 to be associated with an aggressive phenotype in many human cancers.^{14,25,26} MMP-7 has broad substrate specificity against the components of the extracellular matrix.²⁷ Furthermore, MMP-7 plays an important role in the ectodomain shedding of cell-surface molecules, such as E-cadherin.²⁸ Therefore, MMP-7 is considered to be involved in the regulation of these bioactive substances.

In conclusion, this study evaluating NSCLC indicated that the intratumoural Wnt1 expression affects both tumour proliferation and angiogenesis through the induction of its targets such as c-Myc, Cyclin D1, VEGF-A and MMP-7. Therefore, the Wnt1 overexpression can produce more

aggressive malignant tumours during the progression of NSCLCs. Furthermore, the Wnt1 overexpression was an independent prognostic factor for NSCLC patients. To our knowledge, this study is the first comprehensive clinical study demonstrating the clinical significance of the Wnt1 expression in NSCLC. In contrast, no significant difference was observed in the patient survival according to chemotherapy amongst stages II–III NSCLCs in this study. This result might be due to the relatively small number of patients we studied.

The activation of the canonical Wnt/ β -catenin signalling pathway has been shown to play an important role in the development of colorectal carcinoma, which is mainly caused by inactivating mutations of the *adenomatous polyposis coli* (APC) gene or by activating mutations of the β -catenin gene.²⁹ However, previous clinical studies have revealed these mutations to be rare in NSCLCs.¹³ This study has indicated that the Wnt1 overexpression is a possible mechanism of the activation of the canonical Wnt/ β -catenin signalling pathway. Experimental studies demonstrated the Wnt1 expression to be regulated by various molecules, such as NF-kappaB.³⁰ Therefore, the Wnt1 expression might be secondarily regulated in response to a range of changes in many biological molecules during the tumour progression of NSCLC. Further studies should be performed to clarify the mechanism of the Wnt1 overexpression.

From the results given in this study, the Wnt1 can be a candidate of molecular-target therapy for NSCLC. New strategies, such as the RNA inhibition of Wnt1, may be potentially effective treatments for patients with Wnt1-positive NSCLCs.³¹ Because the canonical Wnt1/ β -catenin signalling pathway affects the biological functions via autocrine or paracrine routes, these Wnt1-inhibiting therapies could have bystander effects on tumour tissues. Further studies should be conducted to develop new treatment modalities for Wnt1-positive tumours.

Conflict of interest statement

None declared.

Acknowledgement

This work was supported by Grants-in-Aid for Scientific Research from the Japanese Society for the Promotion of Science, Grant No. 18390379 (C.H.).

REFERENCES

1. Huang C, Liu D, Masuya D, et al. Clinical application of biological markers for treatments of resectable non-small-cell lung cancers. *Br J Cancer* 2005;**92**:1231–9.
2. Dale TC. Signal transduction by the Wnt family of ligands. *Biochem J* 1998;**329**:209–23.
3. You Z, Saims D, Chen S, et al. Wnt signaling promotes oncogenic transformation by inhibiting c-Myc-induced apoptosis. *J Cell Biol* 2002;**157**:429–40.

4. Miller JR, Hocking AM, Brown JD, Moon RT. Mechanism and function of signal transduction by the Wnt/ β -catenin and Wnt/ Ca^{2+} pathways. *Oncogene* 1999;18:7860–72.
5. Mizushima T, Nakagawa H, Kamberov YG, Wilder EL, Klein PS, Rustgi AK. Wnt-1 but not epidermal growth factor induces β -catenin/T-cell factor-dependent transcription in esophageal cancer cells. *Cancer Res* 2002;62:277–82.
6. Korinek V, Barker N, Morin PJ, et al. Constitutive transcriptional activation by a β -catenin-Tcf complex in APC^{-/-} colon carcinoma. *Science* 1997;275:1784–7.
7. He TC, Sparks AB, Rago C, et al. Identification of c-MYC as a target of the APC pathway. *Science* 1998;281:1509–12.
8. Tetsu O, McCormick F. β -catenin regulates expression of cyclin D1 in colon carcinomas cells. *Nature* 1999;398:422–6.
9. Zhang X, Gaspard JP, Chung DC. Regulation of vascular endothelial growth factor by the Wnt and K-ras pathways in colonic neoplasia. *Cancer Res* 2001;61:6050–4.
10. Brabletz T, Jung A, Dag S, Hlubek F, Kirchner T. β -catenin regulates the expression of the matrix metalloproteinase-7 in human colorectal cancer. *Am J Pathol* 1999;155:1033–8.
11. Chen G, Shukeir N, Potti A, et al. Up-regulation of Wnt-1 and β -catenin production in patients with advanced metastatic prostate carcinoma: potential pathogenetic and prognostic implications. *Cancer* 2004;101:1345–56.
12. Zhang WM, Lo Muzio L, Rubini C, Yan G. Effect of WNT-1 on β -catenin expression and its relation to Ki-67 and tumor differentiation in oral squamous cell carcinoma. *Oncol Rep* 2005;13:1095–9.
13. Kotsinas A, Evangelou K, Zacharatos P, Kittas C, Gorgoulis VG. Proliferation, but not apoptosis, is associated with distinct β -catenin expression patterns in non-small-cell lung carcinomas. *Am J Pathol* 2002;161:1619–34.
14. Liu D, Nakano J, Ishikawa S, et al. Overexpression of matrix metalloproteinase-7 (MMP-7) correlates with tumor proliferation, and a poor prognosis in non-small cell lung cancer. *Lung Cancer* 2007;58:384–91.
15. Ben-Ze'ev A, Shtutman M, Zhurinsky J. The integration of cell adhesion with gene expression: the role of β -catenin. *Exp Cell Res* 2000;261:75–82.
16. Dang CV, Resar LM, Emison E, et al. Function of the c-Myc oncogenic transcription factor. *Exp Cell Res* 1999;253:63–77.
17. Volm M, Koomagi R. Prognostic relevance of c-Myc and caspase-3 for patients with non-small cell lung cancer. *Oncol Rep* 2000;7:95–8.
18. Yang G, Timme TL, Frolov A, Wheeler TM, Thompson TC. Combined c-Myc and caveolin-1 expression in human prostate carcinoma predicts prostate carcinoma progression. *Cancer* 2005;103:1186–94.
19. Schauer IE, Siriwardana S, Langan TA, Sclafani RA. Cyclin D1 overexpression vs. retinoblastoma inactivation: implications for growth control evasion in non-small cell and small cell lung cancer. *Proc Natl Acad Sci USA* 1994;91:7827–31.
20. MacLachlan TK, Sang N, Giordano A. Cyclins, cyclin-dependent kinases and cdk inhibitors: implications in cell cycle control and cancer. *Crit Rev Eukaryot Gene Expr* 1995;5:127–56.
21. Caputi M, Groeger AM, Esposito V, et al. Prognostic role of cyclin D1 in lung cancer: relationship to proliferating cell nuclear antigen. *Am J Respir Cell Mol Biol* 1999;20:746–50.
22. Zhang HT, Craft P, Scott PA, et al. Enhancement of tumor growth and vascular density by transfection of vascular endothelial cell growth factor into MCF-7 human breast carcinoma cells. *J Natl Cancer Inst* 1995;87:213–9.
23. Imoto H, Osaki T, Taga S, Ohgami A, Ichiyoshi Y, Yasumoto K. Vascular endothelial growth factor expression in non-small-cell lung cancer: prognostic significance in squamous cell carcinoma. *J Thorac Cardiovasc Surg* 1998;115:1007–14.
24. Johnson DH, Fehrenbacher L, Novotny WF, et al. Randomized phase II trial comparing bevacizumab plus carboplatin and paclitaxel with carboplatin and paclitaxel alone in previously untreated locally advanced or metastatic non-small-cell lung cancer. *J Clin Oncol* 2004;22:2184–91.
25. Yamamoto H, Adachi Y, Itoh F, et al. Association of matrilysin expression with recurrence and poor prognosis in human esophageal squamous cell carcinoma. *Cancer Res* 1999;59:3313–6.
26. Zeng ZS, Shu WP, Cohen AM, Guillem JG. Matrix metalloproteinase-7 expression in colorectal cancer liver metastases: evidence for involvement of MMP-7 activation in human cancer metastases. *Clin Cancer Res* 2002;8:144–8.
27. Wilson CL, Matrisian LM. Matrilysin: an epithelial matrix metalloproteinase with potentially novel functions. *Int J Biochem Cell Biol* 1996;28:123–36.
28. Davies G, Jiang WG, Mason MD. Matrilysin mediates extracellular cleavage of E-cadherin from prostate cancer cells: a key mechanism in hepatocyte growth factor/scatter factor-induced cell-cell dissociation and in vitro invasion. *Clin Cancer Res* 2001;7:3289–97.
29. Bienz M, Clevers H. Linking colorectal cancer to Wnt signaling. *Cell* 2000;103:311–20.
30. Lee TH, Tai DI, Cheng CJ, et al. Enhanced nuclear factor-kappa B-associated Wnt-1 expression in hepatitis B- and C-related hepatocarcinogenesis: identification by functional proteomics. *J Biomed Sci* 2006;13:27–39.
31. You L, He B, Uematsu K, et al. Inhibition of Wnt-1 signaling induces apoptosis in β -catenin-deficient mesothelioma cells. *Cancer Res* 2004;64:3474–8.